

# Ocean Acidification: Monitoring and Mitigation at the Taylor Shellfish Hatchery

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Taylor Shellfish Hatchery  
Quilcene, WA

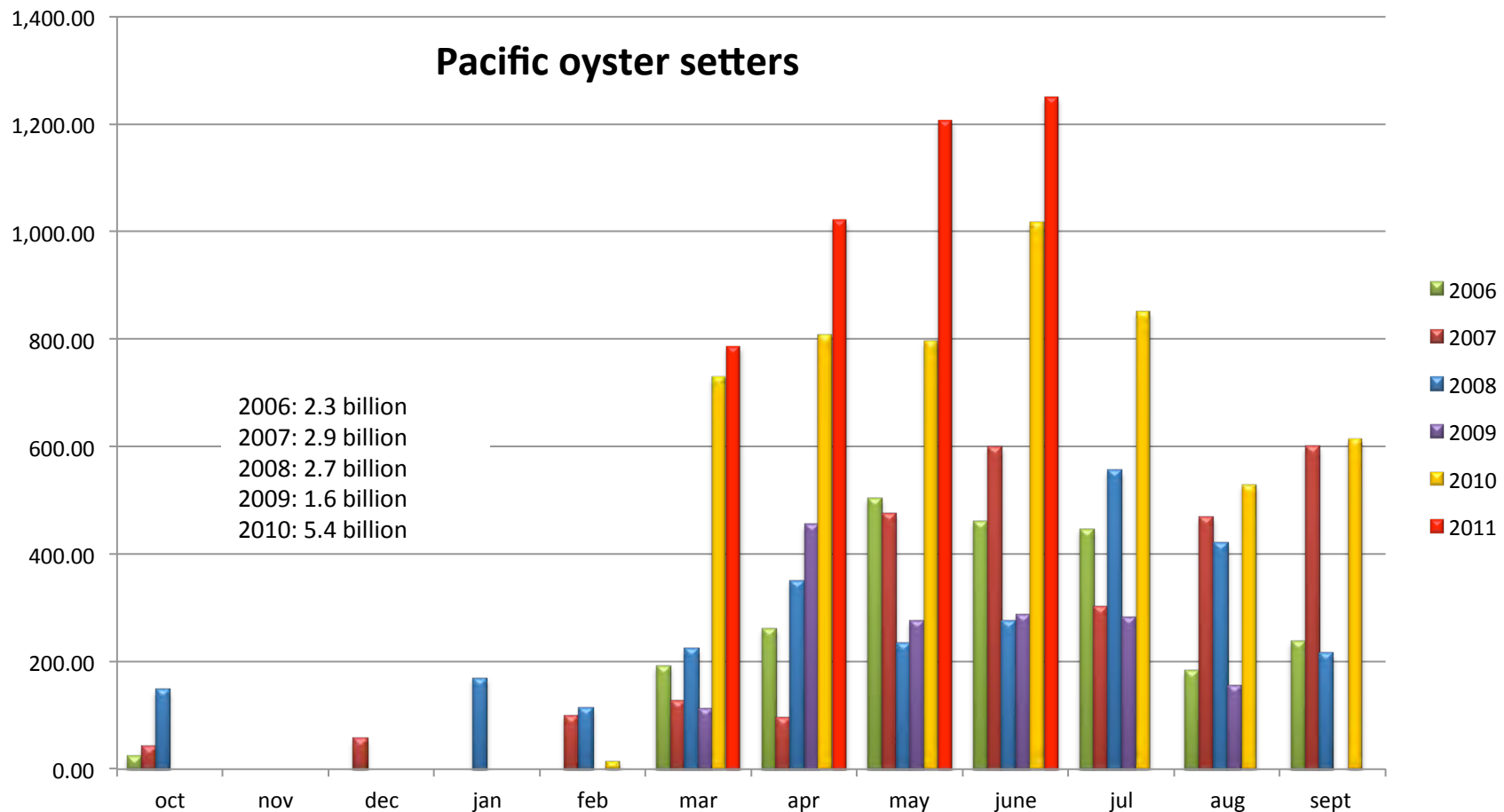


C-CAN Workshop  
July 6-7 2011 Costa Mesa

# Dabob Bay Hatchery



# Pacific Oyster Larvae Production

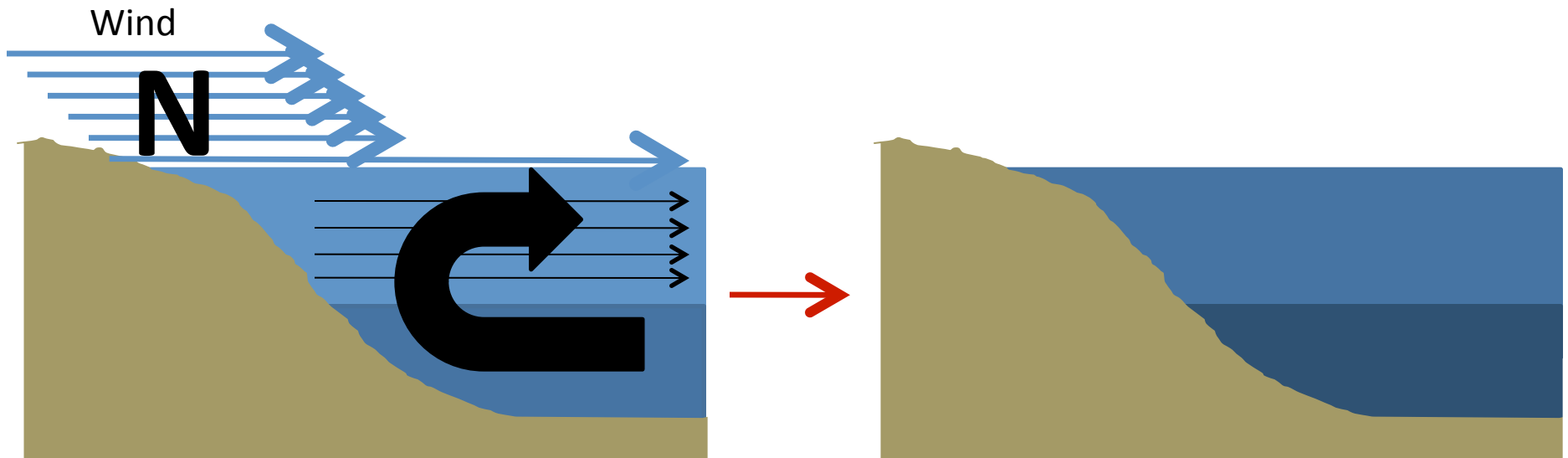


- Production capacity increased from 2006 to 2009 but production decreased
- 2010: Record setting year for oyster larvae production

What is our  
problem?

(We think...)

# Upwelling in Dabob Bay



Combination of 3 low pH/High  $\text{PCO}_2$  sources

- Upwelling in Dabob Bay
- Upwelled ocean water
- Estuarian/ run-off water

Shallow Water

Deep Water

Deeper Water

# Hatchery water supply

- **Shallow intake:** 5 to 15 feet deep, floating  
Shallow water used to raise oyster larvae
- **Deep intake:** 100 feet deep, 4 feet from bottom  
Deep water used for algae production and geoducks

# Shallow and Deep water



# PCO2 Deep/Shallow water





# Deep Water Characteristics

- High  $\text{PCO}_2$
- Low aragonite saturation
- Low pH
- High Salinity
- Low Temperatures
- Reduced chemistry?
- What else?

# Monitoring

# Levels of Monitoring

- Water quality
  - Continuous (Probes, PCO<sub>2</sub>...)
  - Weekly (DIC, nutrients...)
- Larvae performance
- Bacteriology
- Weather



# Water Quality



- YSI 6600 (2) monitoring Shallow and Deep Intakes  
pH, Temperature, Salinity, Oxygen, ORP, Depth (Tide), Chlorophyll, Turbidity.

- YSI 5200A (4) monitoring intakes, water treatments and Bio-assays.  
pH, Temperature, Salinity, Oxygen, ORP

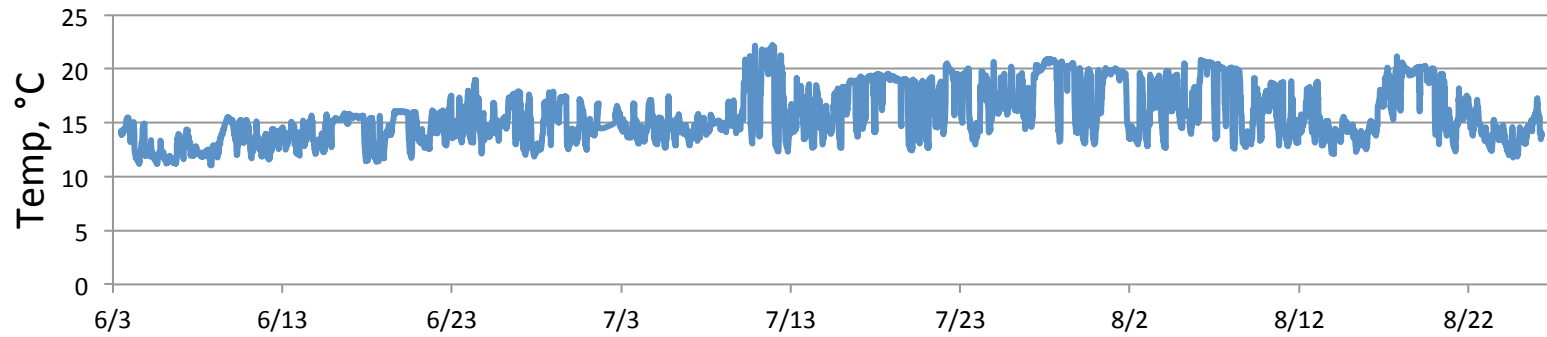


- PCO<sub>2</sub> Monitor (OSU, Burke Hales)  
Continuous monitoring of the PCO<sub>2</sub> in the Shallow intake

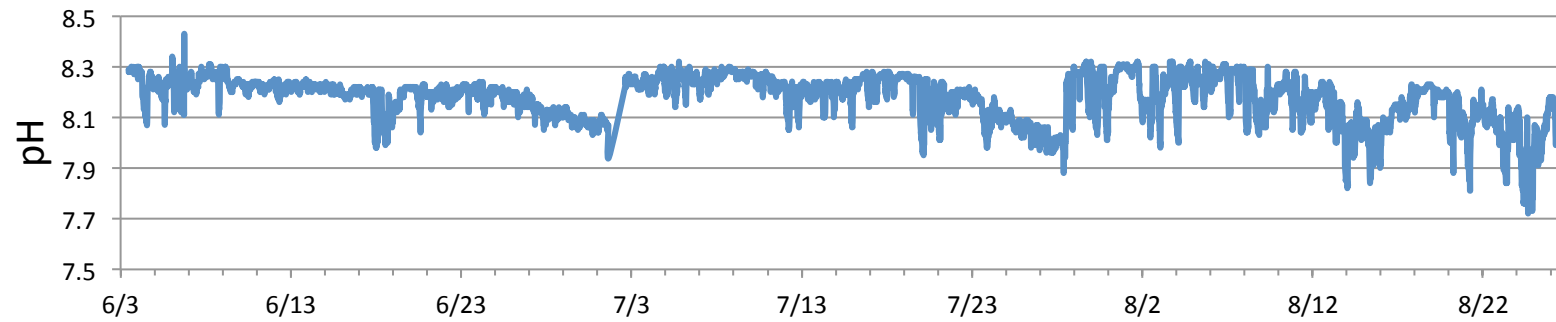


# Continuous Monitoring

## Temperature

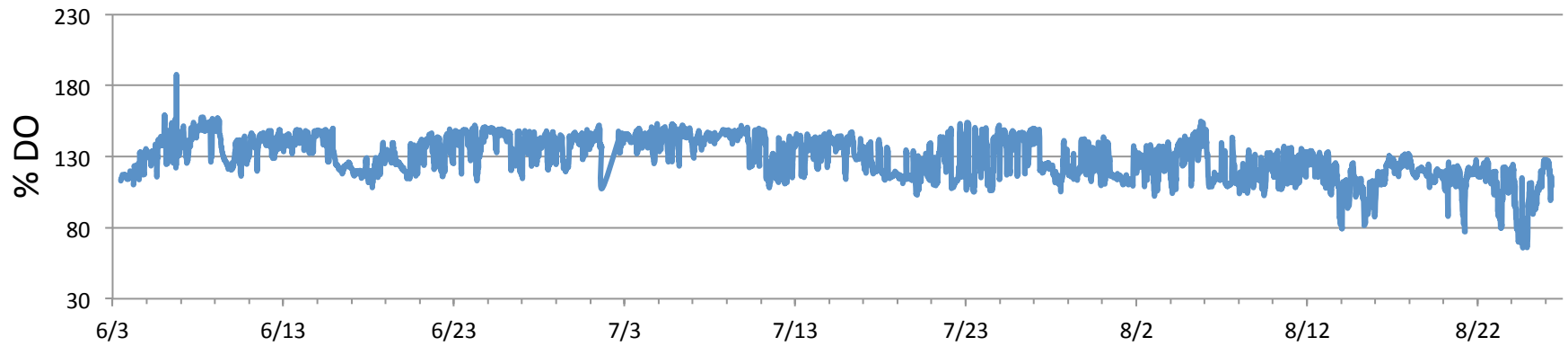


## pH

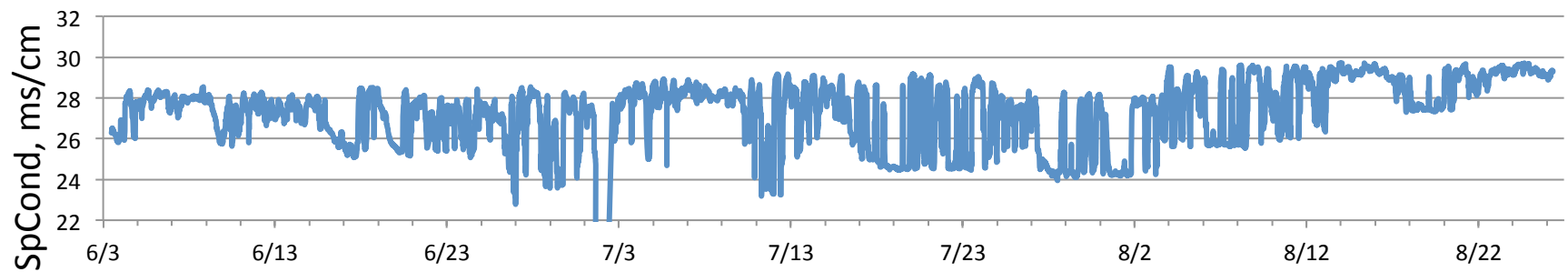


# Continuous Monitoring

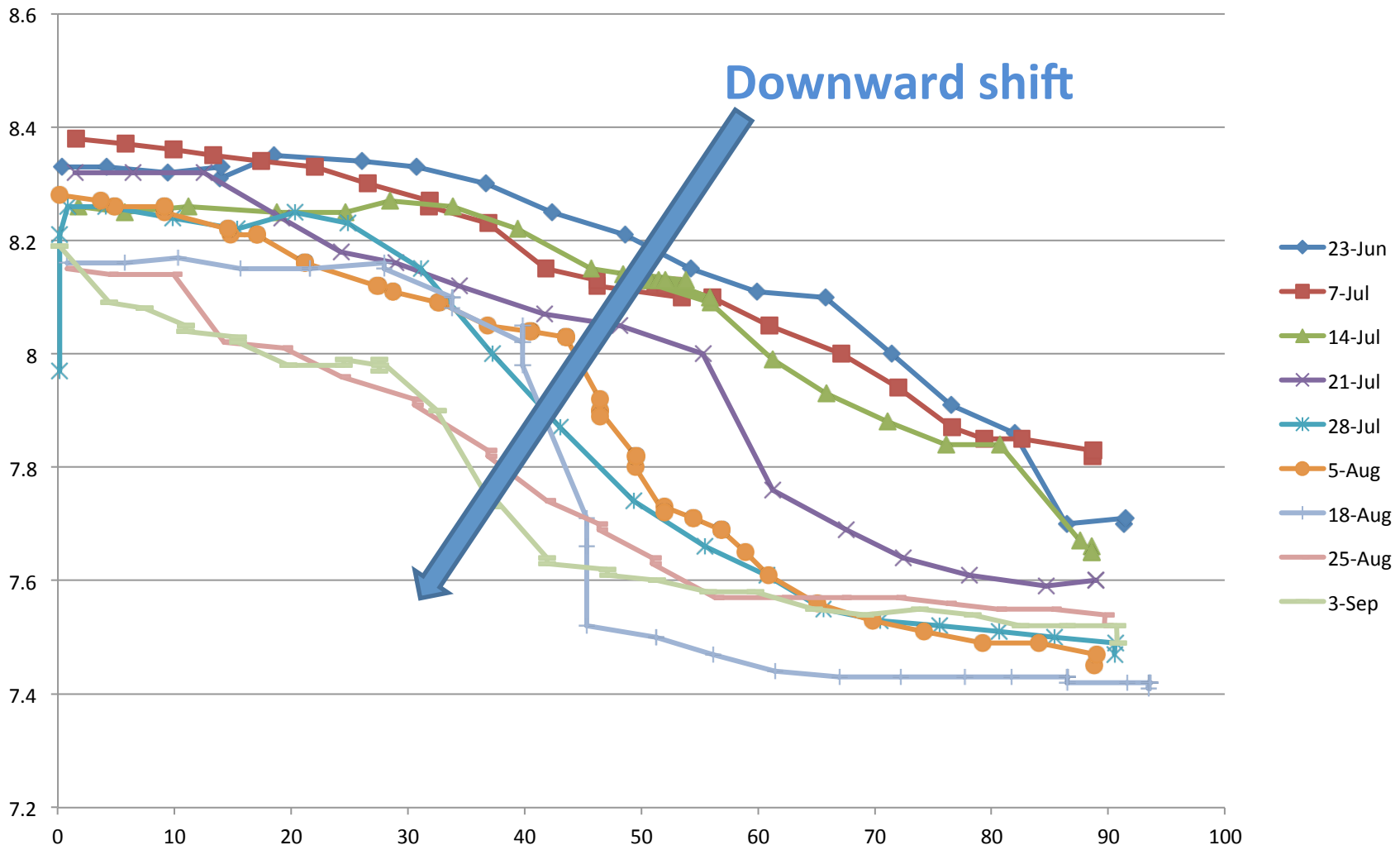
## DO (% saturation)



## Salinity



# Dabob Bay pH Profiles



# Tracking Larvae Performance

## Taylor Shellfish Hatchery larvae spawning sheet

### Spawn information:

Spawn ID	
Tracking number	
Spawn date	
Ploidy	
Spawn type	
Male family	
Female family	
Number of males	
Number of females	
Total number eggs	
Number of eggs kept	
fertilization rate	
Number Dlarvae(CC)	
Number D larvae(DW)	
Number D larvae kept	

### Spawning Water Chemistry:

pH	
O2 mg/l	
O2 (%)	
Temp	
ORP	
Salinity	
Cond.	
SPC	
Alkalinity	
pCO2	
TCO2	
NO2	
NO3	
NH3	

### Larvae information:

<b>First screening date:</b>	
Number 140's	
Number 120's	
Number 100's	
Number 90's	
Number 80's	

<b>Third harvest date:</b>	
Number Setters 250's	
Number setters 236's	
Number Setters 224's	
Number Setters 200's	

<b>First harvest date:</b>	
Number Setters 250's	
Number setters 236's	
Number Setters 224's	
Number Setters 200's	

<b>Fourth harvest date:</b>	
Number Setters 250's	
Number setters 236's	
Number Setters 224's	
Number Setters 200's	

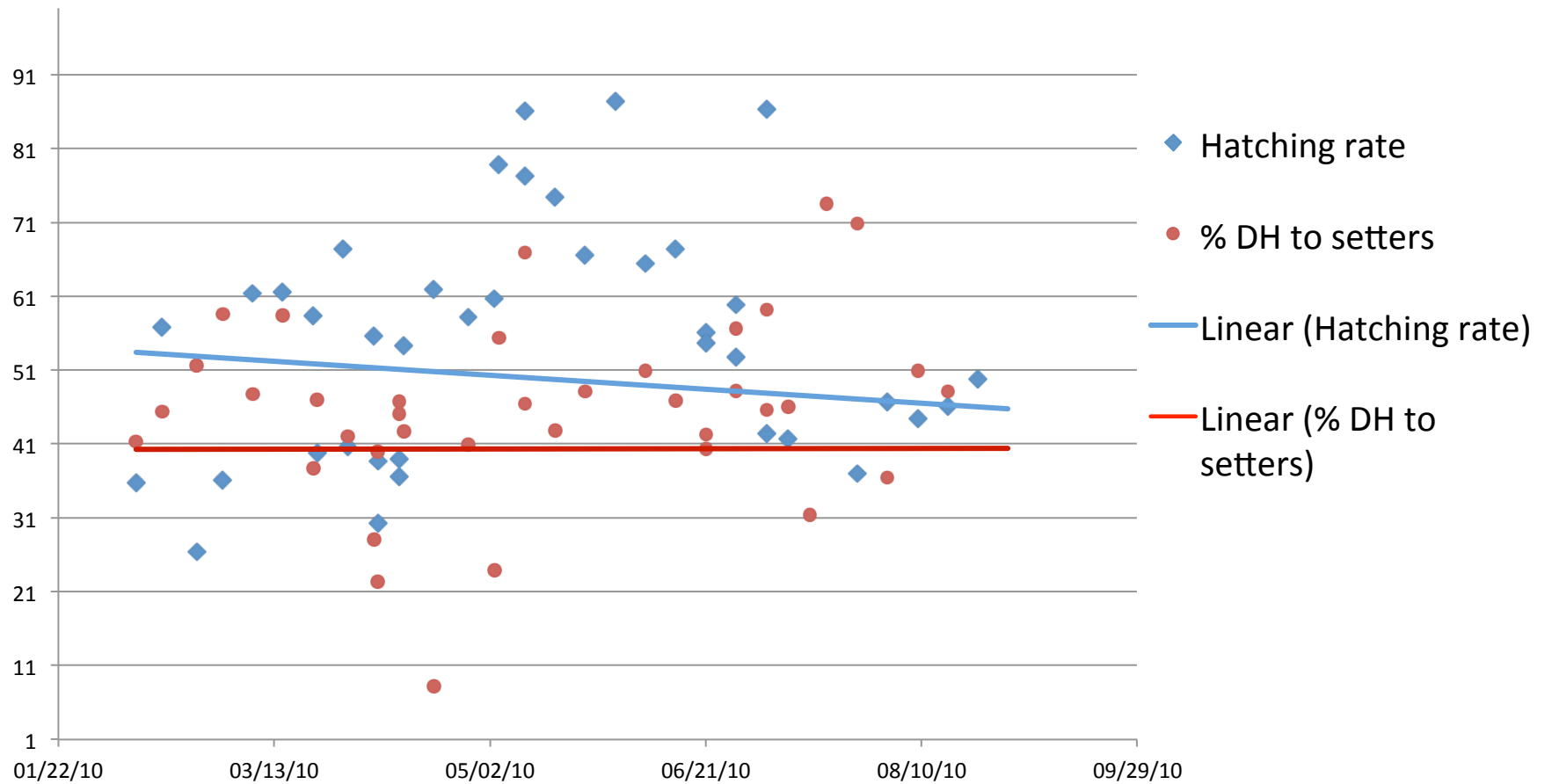
<b>Second harvest date:</b>	
Number Setters 250's	
Number setters 236's	
Number Setters 224's	
Number Setters 200's	

<b>Fifth harvest date:</b>	
Number Setters 250's	
Number setters 236's	
Number Setters 224's	
Number Setters 200's	

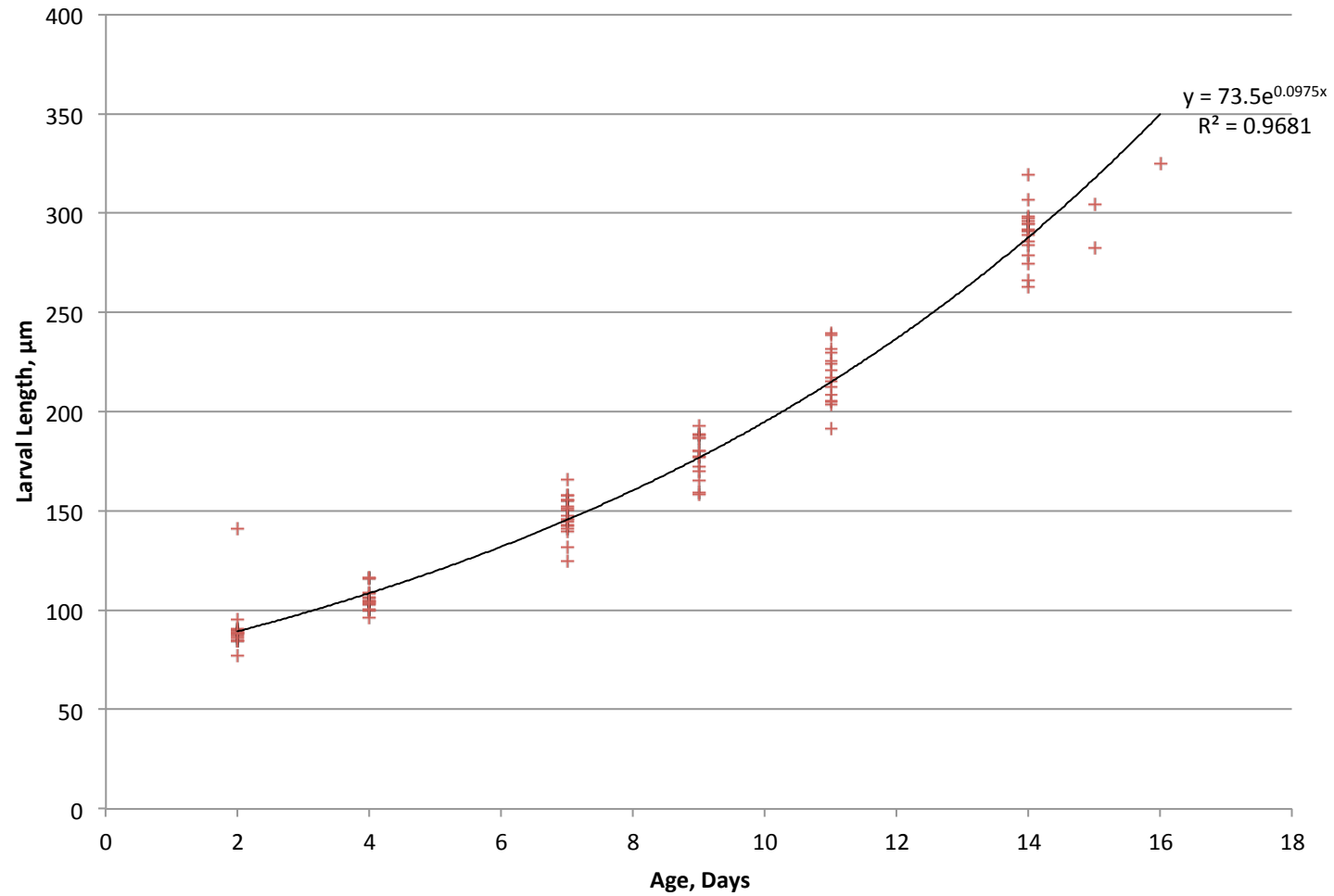
Comments:



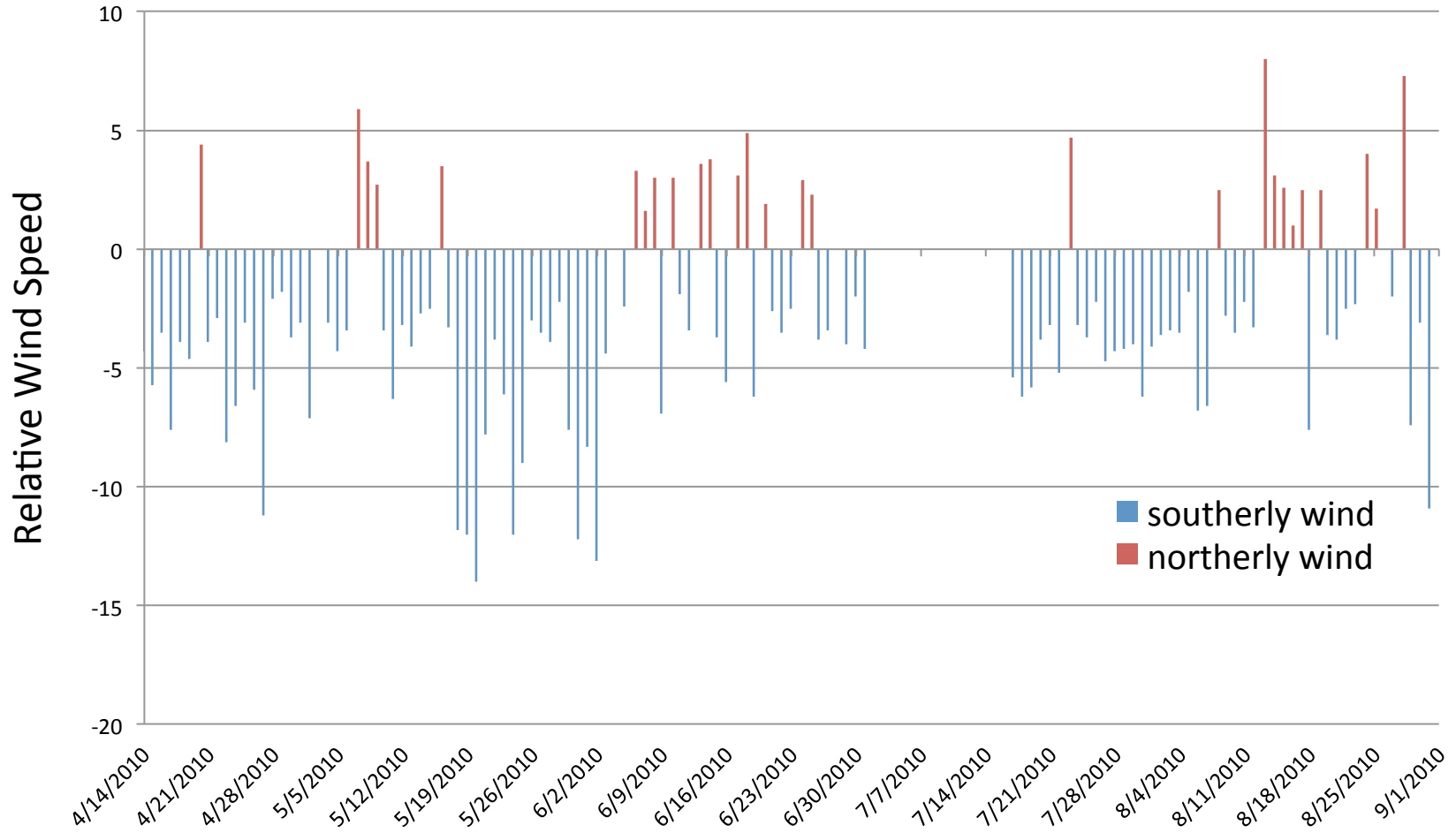
# Larvae Performance



# Larvae growth (16 spawns)



# Wind Direction



# Larvae Bioassays

# Larvae Bioassays

- Based on the comparison of Shallow and Deep water sources.
- Impact of egg hatching condition (0 to 48H PF)
- Impact of larvae rearing conditions
- Comparison of Shallow and Deep water treatments (Sodium carbonate, NaOH, degassing...)

Thank you